Synthesis of Optically Pure ¹³C-Labelled Propionate from Alanine by Asymmetric Hydrolysis using Porcine Kidney Acylase

Katsumi Iida and Masahiro Kajiwara*

Department of Medicinal Chemistry, Meiji College of Pharmacy, 1-22-1 Yato-cho, Tanashi-shi, Tokyo 188, Japan

SUMMARY

Sodium $(R) - [2 - {}^{2}H, 3 - {}^{13}C]$ propionate, sodium $(S) - [2 - {}^{2}H, 3 - {}^{13}C]$ propionate, sodium $(R) - [3 - {}^{13}C] - 2 - fluoropropionate, and sodium <math>(S) - [3 - {}^{13}C] - 2 - fluoropropionate$ were synthesized from $D[(S)] - [3 - {}^{13}C]$ alanine and $L[(R)] - [3 - {}^{13}C]$ alanine, which were obtained by asymmetric hydrolysis using acylase from porcine kidney.

<u>Key words</u>; ¹³C-labelled propionate, deuterium-labelled propionate, fluorine-labelled propionate, ¹³C-labelled alanine, asymmetric hydrolysis.

INTRODUCTION

Recent developments in FT-NMR spectroscopy have allowed stable isotope-labelled positions in compounds to be identified without the need for chemical degradation. This is particularly valuable in studies of biosynthetic pathways. We have already prepared many ¹³C-labelled compounds in our laboratory, and have used them to obtain important information on various biosynthetic processes.¹

0362-4803/91/020201-16\$08.00 © 1991 by John Wiley & Sons, Ltd.

Received August 15, 1990 Revised September 18, 1990 We are currently interested in the stereochemistry of the chain elongation steps in the biosynthesis of macrolide and polyether antibiotics. In order to elucidate this, we required chiral 13 C-labelled propionate double-labelled with deuterium or fluorine. We present here details of the synthesis of these compounds.

RESULTS AND DISCUSSION

$L[(R)] - [3 - {}^{13}C]Alanine (4b) and D[(S)] - [3 - {}^{13}C]alanine (4c)$

¹³C-iodomethane (<u>1</u>) Reaction of with diethyl acetamidomalonate (2) in the presence of sodium in ethyl alcohol gave $[1'-^{13}C]$ diethyl acetamidomethylmalonate (3), which was heated under reflux with concentrated hydrochloric acid to afford $DL-[3-1^3C]$ alanine (4a) in 62 % yield from ^{13}C -iodomethane (1).² Resolution of $DL-[3-1^3C]$ alanine (4a) was carried out by an enzymatic method. DL-[3-¹³C]Alanine (4a) was first heated with acetic acid and acetic anhydride to give DL-[3-¹³C]-N-acetylalanine (<u>5a</u>) in 98 % yield. Asymmetric hydrolysis of $DL-[3-^{13}C]-N$ -acetylalanine (5a) with porcine kidney acylase in 1.5 N ammonium hydroxide gave $L[(R)] - [3 - \frac{13}{C}]$ alanine $(\underline{4b})$ in 38 % yield and D[(S)]-[3-¹³C]-N-acetylalanine (5c). The $D[(S)]-[3-^{13}C]-N-acetylalanine$ (5c) was hydrolyzed with 2 N hydrochloric acid to gave $D[(S)] - [3 - 1^3C]$ alanine (4c) in 32 % yield from $DL-[3-1^3C]-N-acetylalanine (5a)^{3}$ (Fig. 1). Both $L[(R)]-[3-^{13}C]alanine$ (4b) and $D[(S)]-[3-^{13}C]alanine$ (4c) were obtained in high optical purity (100 % ee) as determined by high-performance liquid chromatography (HPLC) using a CROWNPAK CR (-) column.



Figure 1

Sodium (S)-[2-²H,3-¹³C]propionate ($\underline{7c}$) and sodium (R)-[2-²H,3-¹³C]propionate ($\underline{7b}$)

Before ¹³C-labelled sodium $[2-{}^{2}H]$ propionate was synthesized from L[(R)]-[3-¹³C]alanine (<u>4b</u>) and D[(S)]-[3-¹³C]alanine (<u>4c</u>), the optical purity and the absolute configuration of ¹³C-unlabelled sodium $[2-{}^{2}H]$ propionate, synthesized from D[(R)]-alanine and L[(S)]-alanine, were determined by NMR spectroscopy.

Bromination of L[(S)]-alanine (4e) (>98 % ee) with potassium bromide and sodium nitrite in 2.5 N sulfuric acid afforded (S)-2-bromopropionic acid (6e) with retention of configuration in % yield (>98 % ee, the optical purity and absolute 60 were determined by optical rotation; configuration see experimental section). (S)-2-Bromopropionic acid (6e) was heated under reflux with 1 M lithium triethyldeuterioborate in tetrahydrofuran (Superdeuteride; Aldrich) to give sodium $[2-^{2}H]$ propionate (7f) in 81 % yield⁴) (Fig. 2). Compounds (7e) and (7d) were similarly prepared from D[(R)]-alanine (>98 % ee) and DL-alanine, respectively.



Figure 2

Esterification of each sodium $[2-^{2}H]$ propionate (7d, 7e and 7f), derived from DL-, D-, and L-alanine, respectively, with (R)-methyl mandelate (8) using 4-dimethylaminopyridine (DMAP) and dicyclohexylcarbodiimide (DCC) gave the corresponding methyl mandelate derivatives in 47 % yield (9d, <u>9e</u> and <u>9f</u>, respectivery). The optical purity and the absolute configuration were determined by ²H-NMR spectroscopy with broad-band proton decoupling^{4d,5)} (Fig. 3). It was established that sodium (R)-[2-²H]propionate ($\frac{7f}{1}$) (>97 % ee) had been formed from L[(S)]-alanine (<u>4e</u>) (>98 % ee), and sodium (S)-[2-²H]propionate $(\underline{7e})$ (>97 % ee) from D[(R)]-alanine (>98 %ee). Futhermore, we applied the circular dichroic (CD) spectra to the determination of the sense of chirality of the chiral sodium $[2-^{2}H]$ propionate (7e and 7f). The CD spectra are shown in Fig. 4, where $(S)-[2-^{2}H]$ propionate (7e) and $(R)-[2-^{2}H]$ propionate (7f) show the positive curve and negative curve respectively. These data also indicate that the optical purity of <u>7e</u> and <u>7f</u> are numerically almost equal to those described above NMR method.

On the basis of the results , sodium $(S)-[3-^{13}C,2-^{2}H]$ propionate $(\underline{7c})$ (>97 % ee) was prepared via $(R)-[3-^{13}C]-2$ -bromopropionic acid ($\underline{6b}$) obtained from $L[(R)]-[3-^{13}C]$ alanine ($\underline{4b}$), and sodium (R)- $[3-^{13}C,2-^{2}H]$ propionate ($\underline{7b}$) (>97 % ee) was similarly prepared via $(S)-[3-^{13}C]-2$ -bromopropionic acid ($\underline{6c}$) obtained from $D[(S)]-[3-^{13}C]$ alanine ($\underline{4c}$) (Fig. 5).





Figure 4



Sodium $(R)-[3-^{13}C]-2-fluoropropionate (<u>10b</u>) and sodium (S) [3-^{13}C]-2-fluoropropionate (<u>10c</u>)$

In the same way, the optical purity of ¹³C-unlabelled sodium 2-fluoropropionate, synthesized from D- and L-alanine, was determined by NMR. With retention of configuration as aforesaid



bromination, L[(S)]-alanine (<u>4e</u>) was treated with sodium nitrite hydrogen fluoride-pyridine to in give sodium (S)-2fluoropropionate (10e) in 95 % yield⁶⁾ (Fig. 6). Compounds (<u>10f</u>) and (10d) were similarly prepared from D- and DL-alanine. Esterification of each sodium 2-fluoropropionate (10d, 10e and 10f) with (R)-methyl mandelate (8) gave the corresponding methyl mandelate derivative in 49 % yield (11d, 11e and 11f, respectively). The optical purity were determined by 1 H-NMR spectroscopy (Fig. 7) and CD spectra (Fig. 8). It was established that sodium (S)-2-fluoropropionate (10e) (>93 % ee) had been formed from L[(S)]-alanine (4e) (>98 % ee) and sodium (R)-2-fluoropropionate (10f) (>93 % ee) from D[(R)]-alanine (>98 % ee).







the On the basis of above results, sodium $(R)-[3-^{13}C]-2-fluoropropionate (10b)$ (>93 % ee) was prepared from $L[(R)] - [3 - 1^{3}C]alanine$ $(s) - [3 - {}^{13}C] -$ (<u>4b</u>), and sodium 2-fluoropropionate (10c) (>93 % ee) was similarly prepared from $D[(S)] - [3 - {}^{13}C]alanine (4c) (Fig. 9).$





EXPERIMENTAL

Materials

 13 C-Iodomethane (99 atom % 13 C) was supplied by Cambridge

Isotope Laboratories. Lithium triethyldeuterioborate (1 M in tetrahydrofuran) (Superdeuteride) and hydrogen fluoride-pyridine were obtained from Aldrich. D-Alanine (>98 % ee) and L-alanine (>98 % ee) were obtained from Tokyo Kasei Kogyo. (R)-Methyl mandelate (99 % ee) was prepared from (R)-mandelic acid (99 %ee), which was obtained from Merck. Other reagents were obtained from Tokyo Kasei Kogyo and Wako. Acylase from porcine kidney was purchased from Sigma and had a specific activity of 1580 units per mg of protein. Water was ion-exchanged and distilled before use.

Instruments

Melting point (m.p.) determinations were carried on a Yazawa micro melting point apparatus, Type BY-1; values are uncorrected. Infrared spectra (IR) were recorded on a JASCO DS-801 spectrometer, and positions of absorption maxima are reported in reciprocal centimeters. ¹H-NMR spectra were recorded on a JEOL GSX-400 spectrometer in CDCl₃ solution with tetramethylsilane (TMS) as an internal standard, or in D_2O solution referenced to the solvent peak. ¹³C-NMR spectra were recorded on a JEOL

GSX-400 (100 MHz) spectrometer in CDCl, solution referenced to D_2O solution with solvent or in the peak, sodium 3-trimethylsilylpropionate (TSP) as an internal standard. ²H-NMR spectra were recorded on a JEOL GSX-270 (41.3 MHz) spectrometer in CDCl₃ solution referenced to the solvent peak, with broad-band proton decoupling. Fast atom bombardment mass spectra (FAB-MS) were obtained on a JEOL JMS-DX302. CD spectra were recored on a JASCO J-500C spectrometer. The HPLC system consisted of a JASCO 880-PU pump and a JASCO 870-UV detector; column, CROWNPAK CR (-), 15 cm X 0.4 cm I.D. (Daicel); column temperature, 0 °C; mobile phase, aqueous HClO, pH 1.0; flow rate, 0.4 ml/min; UV detection wavelength, 200 nm. The retention times observed were 5.6 min $L[(R)] - [3 - {}^{13}C]alanine$ (4b), for and 12.6 min for $D[(S)]-[3-^{13}C]$ alanine (4c). Optical rotation were recorded on a JASCO DIP-360 polarimeter.

$[1'-^{13}C]$ Diethyl acetamidomethylmalonate $(\underline{3})$

A solution of sodium (0.85 g, 37.0 mmol) in dry ethyl alcohol (16 ml) was added to a solution of diethyl acetamidomalonate (2) (7.65 g, 35.2 mmol) in dry ethyl alcohol (19 ml) at 0 °C under argon, and the mixture was stirred for 10 min at room temperature. To this solution, ¹³C-iodomethane (<u>1</u>) (5.0 g, 35.0 mmol) was added dropwise at 0 °C, and the whole was stirred for 5 min at this temperature, then heated under reflux for 15 hr. The reaction was quenched with water (10 ml) and the mixture was extracted with ether (50 X 8 ml). The combined extracts were washed with brine (200 ml), dried over magnesium sulfate and evaporated. Chromatography of the crude product on silica gel with hexane:ether (1:1) gave $[1'-^{13}C]$ diethyl acetamidomethylmalonate (<u>3</u>) (5.50 g, 68 %), m.p. 88.0 \sim 89.8 °C; ¹H-NMR (CDCl₃) 1.26 (t, 6H, J_{HH}=7.2 Hz, CH₃CH₂), 1.75 (d, 3H, J_{13CH}=131.9 Hz, ¹³CH₃C), 2.02 (s, 3H, CH₃CO), 4.25 (q, 4H,

 $J_{HH} = 7.2 \text{ Hz}, \text{ CH}_3 \text{CH}_2$; ¹³C-NMR (CDCl₃) 20.8; FAB-MS m/z 233 (M⁺+1, 100 %); IR (CHCl₃) 3425 (NH), 1735 (COOC), 1685 (NC).

$DL-[3-^{13}C]$ Alanine (<u>4a</u>)

A solution of $[1'-^{13}C]$ diethyl acetamidomethylmalonate (<u>3</u>) (5.50 g, 23.7 mmol) in concentrated hydrochloric acid (25 ml) was heated under reflux for 2 hr. The reaction mixture was cooled to room temperature and evaporated. The residue was dissolved in a minimum volume of 50 % ethyl alcohol. Pyridine was added to this solution until crystallization occurred, and the crystals were collected by centrifugation at 1400 G for 15 min and washed with ethyl alcohol. Recrystallization from water-ethyl alcohol gave DL-[3-¹³C]alanine (<u>4a</u>) (1.94 g, 91 %), m.p. 292 \sim 294 °C; ¹H-NMR (D₂O) 1.38 (dd, 3H, J_{HH}=7.2 Hz, J_{13CH}=129.6 Hz, ¹³CH₃CH), 3.68 (dq, 1H, J_{HH}=7.2 Hz, J_{13CCH}=4.6 Hz, ¹³CH₃CH); ¹³C-NMR (D₂O) 18.9; FAB-MS m/z 91 (M⁺+1, 34 %); IR (KBr) 3082 (NH), 1685, 1590 (COOH).

$DL-[3-^{13}C]-N-Acetylalanine (5a)$

A suspension of $DL-[3-^{13}C]alanine$ (<u>4a</u>) (2.34 g, 26.0 mmol) in acetic acid (11 ml) was heated to 100 °C for 5 hr. Acetic anhydride (3.5 ml) was then added and the mixture was heated to 110 °C until a solution was formed. This temperature was held for 5 min, then the reaction mixture was quenched with water (50 ml) and evaporated. Recrystallization from water-acetone gave $DL-[3-^{13}C]$ -N-acetylalanine (<u>5a</u>) (3.36 g, 98 %), m.p. 123 \sim 125 °C; ¹H-NMR (D₂O) 1.31 (dd, 3H, J_{HH}=7.3 Hz, J_{13CH}=130.3 Hz, ¹³CH₃CH), 1.92 (s, 3H, CH₃CO), 4.23 (m, 1H, ¹³CH₃CH); ¹³C-NMR (D₂O) 20.2; FAB-MS m/z 133 (M⁺+1, 100 %); IR (KBr) 3330 (NH), 1720 (CO), 1590 (NC).

$L[(R)]-[3-^{13}C]Alanine$ (4b) and $D[(S)]-[3-^{13}C]Alanine$ (4c)

Porcine kidney acylase (158 mg, 250 K units) was added to a solution (pH 8.0) of $DL-[3-^{13}C]-N$ -acetylalanine (5a) (2.77 g, 21.0 mmol) in 1.5 N ammonium hydroxide (18 ml), and the mixture was stirred at 38 °C. After 23 hr, the reaction mixture was boiled for a few minutes, and the denatured enzyme was removed by centrifugation at 1400 G for 15 min. The supernatant solution was concentrated, ethyl alcohol (5 ml) was added, and the resulting crystals were separated from the solution, collected by centrifugation at 1400 G for 15 min, and washed with ethyl alcohol (5 ml). The process was repeated twice, and finally recrystallization from water-ethyl alcohol gave $L[(R)]-[3-^{13}C]alanine (4b)$ (721 mg, 38 %). After isolation of crude $L[(R)] - [3 - 1^{3}C]$ alanine (4b), the supernatant solution was evaporated. The residue was subjected to column chromatography with on Dowex 50W X – 4 and elution water afforded $D[(S)] - [3 - {}^{13}C] - N$ -acetylalanine (5c) (1.28 g, 46 %). A solution of $D[(S)]-[3-^{13}C]-N-acetylalanine (5c) (1.28 g, 9.70 mmol) in 2 N$ hydrochloric acid (20 ml) was heated under reflux for 4 hr, then cooled to room teperature and evaporated. The residue was dissolved in a minimum volume of 50 % ethyl alcohol. Pyridine was added to this solution until crystallization occurred, and the crystals were collected by centrifugation at 1400 G for 15 min and washed with ethyl alcohol. Recrystallization from water-ethyl alcohol gave $D[(S)]-[3-^{13}C]$ alanine (4c) (605 mg, 32 %, 2 steps). Analytical data of 4b and 4c were exactly harmonized with $DL-[3-^{13}C]$ alanine (4a) and no distinguished between 4b and <u>4c</u>.

(R)-[3-¹³C]-2-Bromopropionic acid ($\underline{6b}$) and (S)-[3-¹³C]-2-Bromopropionic acid ($\underline{6c}$)

Sodium nitrite (1.0 g, 14.5 mmol) was added to a solution of

L[(R)]-[3-¹³C]alanine (<u>4b</u>) (404 mg, 4.48 mmol) and potassium bromide (3.5 g, 29.4 mmol) in 2.5 N sulfuric acid (37 ml) at -10 °C, and the reaction mixture was stirred for 3 hr, then extracted with ether (150 X 3 ml). The combined extracts were washed with brine (200 ml), dried over magnesium sulfate and evaporated. Distillation gave (R)-[3-¹³C]-2-bromopropionic acid (<u>6b</u>) (411 mg, 60 %); b.p. 95 °C (18 mmHg), ¹H-NMR (CDCl₃) 1.85 (dd, 3H, J_{HH}=6.9 Hz, J_{13CH}=131.0 Hz, ¹³CH₃CHBr), 4.41 (dq, 1H, J_{HH}=6.9 Hz, J_{13CCH}=2.6 Hz, ¹³CH₃CHBr); ¹³C-NMR (CDCl₃) 21.4; FAB-MS m/z 154 (M⁺+1, 4 %), 156 (M⁺+3, 4 %); IR (CHCl₃) 1725 (CO).

2-Bromopropionic acid, ¹H-NMR (CDCl₃) 1.85 (d, 3H, $J_{HH}=6.9$ Hz, CH_3CHBr), 4.41 (q, 1H, $J_{HH}=6.9$ Hz, CH_3CHBr).

(S)-2-Bromopropionic acid (<u>6e</u>) prepared from L[(S)]-alanine (<u>4e</u>), $[\alpha]_{D}^{23}$ =-26.2 (c=15.1, CHCl₃), Lit; $[\alpha]_{D}^{25}$ =-25.3 (c=15.0, CHCl₃)⁷⁾.

(R)-2-Bromopropionic acid prepared from D[(R)]-alanine, [α]²³_D=26.6 (c=15.1, CHCl₃).

Sodium $(S) - [2 - {}^{2}H, 3 - {}^{13}C]$ propionate $(\underline{7c})$ and Sodium $(R) - [2 - {}^{2}H, 3 - {}^{13}C]$ propionate $(\underline{7b})$

(R)-[3-¹³C]-2-Bromopropionic acid (<u>6b</u>) (411 mg, 2.67 mmol) was added to 1 M lithium triethyldeuterioborate in tetrahydrofuran (5 ml) at 0 °C under argon. This mixture was heated under reflux for 4 hr, then the reaction was quenched with water (10 ml), and the solution was adjusted to pH 4.0 with concentrated sulfuric acid. Leaving a minimum volume of the solvent, the propionate was distilled off with water. Water was added to the residue, and distillation was continued. The distillate was combined, and adjusted to pH 8.0 with a 0.1 N aqueous sodium carbonate solution. Freeze-drying gave sodium (S)-[2-²H,3-¹³C]propionate (7<u>c</u>) (211 mg, 81 %), ¹H-NMR (D₂O) 0.93 (dd, 3H, J_{HH} =7.6 Hz, J_{13CH} =127.2 Hz, ¹³CH₃CHD), 2.05 (m, 1H, ¹³CH₃CHD); ¹³C-NMR (D₂O) 12.9. Sodium $[2-^{2}H]$ propionate $(\underline{7d}, \underline{7e} \text{ and } \underline{7f}), {}^{1}H-NMR (D_{2}O) 0.96 (d, 3H, J_{HH}=7.7 Hz, CH_{3}CHD), 2.07 (tq, 1H, J_{HH}=7.7 Hz, J_{HD}=2.3 Hz, CH_{3}CHD).$

Sodium $(R)-[3-^{13}C]-2-fluoropropionate (10b)$ and Sodium $(S)-[3-^{13}C]-2-fluoropropionate (10c)$

Sodium nitrite (260 mg, 3.77 mmol) was added to a solution of $L[(R)] - [3 - 1^{3}C]$ alanine (<u>4b</u>) (99 mg, 1.10 mmol) in hydrogen fluoride-pyridine (2 ml) at -10 °C. The reaction mixture was stirred for 3 hr at room temperature, then the reaction was quenched with water (10 ml). The resulting solution was adjusted to pH 4.0 with a sodium hydrogen carbonate solid. Leaving a minimum volume of the solvent, the propionate was distilled off with water. Water was added to the residue and distillation was continued. The distillate was combined, and adjusted to pH 8.0 with a 0.1 N aqueous sodium carbonate solution. Freeze-drying gave sodium (R)- $[3-^{13}C]-2-fluoropropionate (<u>10b</u>) (119 mg, 95 %),$ ¹H-NMR ($D_{2}O$) 1.40 (ddd, 3H, $J_{HH}=6.9$ Hz, $J_{HF}=24.4$ Hz, $J_{13CH}=128.6$ Hz, ${}^{13}CH_{3}CHF$), 4.83 (ddg, 1H, $J_{HH}=6.9$ Hz, $J_{HF}=51.1$ Hz, $J_{13CCH}=2.8$ Hz, 13 CH₃C<u>H</u>F); 13 C-NMR (D₂O) 21.2 (d, J_{HF}=22.0 Hz). Sodium 2-fluoropropionate (10d, 10e and 10f), ¹H-NMR (D₂O) 1.41 (dd, 3H, $J_{HH}=6.9$ Hz, $J_{HF}=24.4$ Hz, $C\underline{H}_{3}CHF$), 4.84 (dq, 1H, $J_{HH}=6.9$ Hz, $J_{HF} = 50.8$ Hz, CH_3CHF).

(R)-Methyl mandelate derivatives (<u>9d</u>, <u>9e</u>, <u>9f</u>, <u>11d</u>, <u>11e</u> and <u>11f</u>) of 13 C-unlabelled propionates

A solution of deuterium-labelled sodium propionate (7d) (202 mg, 2.08 mmol) in water was adjusted to pH 4.0 with 1 N hydrochloric acid and extracted with ether. The combined extracts were dried over magnesium sulfate and evaporated. Dicyclohexylcarbodiimide (DCC) (454 mg, 2.20 mmol) and (R)-methyl mandelate (8) [(R)-methyl 2-hydroxy-2-phenylethanoate] (292 mg, 1.76 mmol) were added to a solution of the residue and 4-dimethylaminopyridine (DMAP) (4.7 mg) in dry dichloromethane (10 ml) at -10 $^{\circ}\text{C}$ under argon, and the reaction mixture was stirred for 3 hr at room temperature, then filtered and evaporated. Chromatography of the crude product on silica gel with ether: hexane (1:2) gave the corresponding (R)-methyl mandelate derivative (9d) (185 mg, 47 8) of sodium $[2-^{2}H]$ propionate $(\underline{7d})$.

(R)-Methyl mandelate derivative (<u>9d</u>) of sodium $[2-^{2}H]$ propionate (<u>7d</u>), ¹H-NMR (CDCl₃) 1.20 (d, 3H, J_{HH}=7.4 Hz, CH₃CHD), 2.45, 2.51 (tq, 1H, J_{HH}=7.4 Hz, J_{HD}=2.4 Hz, CH₃CHD), 3.72 (s, 3H, CH₃O), 5.94 (s, 1H, CHO), 7.39, 7.46 (m, 5H, phenyl); ²H-NMR (CDCl₃) 2.47, 2.54 (br, 1D, CHD).

(R)-Methyl mandelate derivative $(\underline{9f})$ of sodium (R)-[2-²H]propionate $(\underline{7f})$, ¹H-NMR (CDCl₃) 1.20 (d, 3H, J_{HH}=7.4 Hz, CH₃CHD), 2.51 (tq, 1H, J_{HH}=7.4 Hz, J_{HD}=2.4 Hz, CH₃CHD), 3.72 (s, 3H, CH₃O), 5.94 (s, 1H, CHO), 7.39, 7.46 (m, 5H, phenyl); ²H-NMR (CDCl₃) 2.47 (br, 1D, CHD).

(R)-Methyl mandelate derivative (<u>9e</u>) of sodium (S)-[$2-^{2}$ H]propionate (<u>7e</u>), ¹H-NMR (CDCl₃) 1.20 (d, 3H, J_{HH}=7.4 Hz, CH₃CHD), 2.45 (tq, 1H, J_{HH}=7.4 Hz, J_{HD}=2.4 Hz, CH₃CHD), 3.72 (s, 3H, CH₃O), 5.94 (s, 1H, CHO), 7.39, 7.46 (m, 5H, phenyl); ²H-NMR (CDCl₃) 2.54 (br, 1D, CHD).

A solution of sodium 2-fluoropropionate (<u>10d</u>) (241 mg, 2.11 mmol) in water was adjusted to pH 4.0 with 1 N hydrochloric acid and extracted with ether. The combined extracts were dried over magnesium sulfate and evaporated. Dicyclohexylcarbodiimide (DCC) (561 mg, 2.72 mmol) and (R)-methyl mandelate (<u>8</u>) [(R)methyl 2-hydroxy-2-phenylethanoate] (291 mg, 1.75 mmol) were added to a solution of the residue and 4-dimethylaminopyridine (DMAP) (4.7 mg) in dry dichloromethane (10 ml) at -10 °C under argon, and the reaction mixture was stirred for 3 hr at room temperature, then

filtered and evaporated. Chromatography of the crude product on silica gel with ether:hexane (1:5) gave the corresponding (R)-methyl mandelate derivative (<u>11d</u>) (206 mg, 49 %) of sodium 2-fluoropropionate (<u>10d</u>).

(R)-Methyl mandelate derivative (<u>11d</u>) of sodium 2-fluoropropionate (<u>10d</u>), ¹H-NMR (CDCl₃) 1.63, 1.71 (dd, 3H, $J_{HH}=6.9$ Hz, $J_{HF}=23.6$ Hz, $CH_{3}CHF$), 3.74 (s, 3H, $CH_{3}O$), 5.14, 5.18 (dq, 1H, $J_{HH}=6.9$ Hz, $J_{HF}=48.4$ Hz, $CH_{3}CHF$), 6.03 (s, 1H, CHO), 7.41, 7.46 (m, 5H, phenyl).

(R)-Methyl mandelate derivative (<u>11e</u>) of sodium (S)-2-fluoropropionate (<u>10e</u>), ¹H-NMR (CDCl₃) 1.63 (dd, 3H, $J_{HH}=6.9$ Hz, $J_{HF}=23.6$ Hz, $C\underline{H}_{3}CHF$), 3.74 (s, 3H, $C\underline{H}_{3}O$), 5.18 (dq, 1H, $J_{HH}=6.9$ Hz, $J_{HF}=48.4$ Hz, $CH_{3}C\underline{H}F$), 6.03 (s, 1H, $C\underline{H}O$), 7.41, 7.46 (m, 5H, phenyl).

(R)-Methyl mandelate derivative (<u>11f</u>) of sodium (R)-2-fluoropropionate (<u>10f</u>), ¹H-NMR (CDCl₃) 1.71 (dd, 3H, $J_{HH}=6.9$ Hz, $J_{HF}=23.6$ Hz, $C_{H_3}CHF$), 3.74 (s, 3H, $C_{H_3}O$), 5.14 (dq, 1H, $J_{HH}=6.9$ Hz, $J_{HF}=48.4$ Hz, CH_3CHF), 6.03 (s, 1H, $C_{H}O$), 7.41, 7.46 (m, 5H, phenyl).

ACKNOWLEDGMENT

We wish to thank Mr. Hiroaki Utsumi of JEOL Co. for measurement of the ²H-NMR spectra and Dr. Nobukazu Fukuhara for discussion of fluorine chemistry. Particular thanks are due to Mr. Takashi Takakuwa in JASCO Co. for measurement and interpretation of the CD spectra.

REFERENCES

a) Kurumaya, K., Okazaki, T., Seido, N., Akasaka, Y.,
 Kawajiri, Y., Kajiwara, M., and Kondo, M. -J. labelled
 Compounds and Radiopharmaceuticals <u>27</u>: 217 (1989)

- b) Kurumaya, K., Okazaki, T. and Kajiwara, M. -Chem. Pharm.Bull. 37: 1151 (1989)
- 2) Albertson, N.F. -J. Amer. Chem. Soc. 68: 450 (1946)
- 3) a) Kerr, V.N. and Ott, D.G. -J. labelled Compounds and Radiopharmaceuticals <u>15</u>: 503 (1978)
 - b) Mayo, D.W., Butcher, S.S. and Pike, R.M. -Microscale Organic Laboratory p.270 John Wiley & Sons, Inc., Canada (1986)
- 4) a) Retey, J., Ronchi, U. and Arigoni, D. -Experientia <u>22</u>:72 (1966)
 - b) Zagalak, B., Frey, P.A., Karabatsos, G.L. and Abeles,
 R.A. -J. Biol. Chem. <u>241</u>: 3028 (1966)
 - c) Prescott, D.J. and Rabinowitz, J.L. -J. Biol. Chem. <u>243</u>: 1551 (1968)
 - d) Sood, G.R., Ashworth, D.M., Ajaz, A.A. and Robinson, J.A.
 -J. Chem. Soc. Perkin Trans. I 3183 (1988)
 - e) Armarego, W.L.F, Milloy, B.A. and Pendergast, W. -J. Chem. Soc. Perkin Trans. I 2229 (1976)
- 5) a) Parker, D. -J. Chem. Soc. Perkin Trans. II 83 (1983)
 - b) Brown, J.M. and Parker, D. -Tetrahedron Lett. <u>22</u>: 2815 (1981)
- 6) Olah, G.A. and Welch, J. -Synthesis 652 (1974)
- 7) Kirchner, G., Scollar, M.P. and Klibanov, A.M. -J. Amer. Chem. Soc. <u>107</u>: 7072 (1985)